Progress Toward a Global Genealogy of Common Carp (Cyprinus carpio L.) Strains Using Mitochondrial Nucleotide Sequences Data

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Abstract

As part of a study of genetic variation in the Vietnamese strains of the common carp (Cyprinus carpio L) using direct DNA sequencing of mitochondrial control and ATPase6/8 gene regions, samples from a number of other countries were analyzed for comparison. Results show that the levels of sequence divergence in common carp is low on a global scale, with the Asian carp having the highest diversity while Koi and European carp are invariant. A genealogical analysis supports a close relationship among Vietnamese, Koi, Chinese Color and, to a lesser extent, European carp. Koi carp appear to have originated from a strain of Chinese red carp. There is considerable scope to extend this research through the analysis of additional samples of carp from around the world, especially from China, in order to generate a comprehensive global genealogy of common carp strains.

Introduction

The sequence divergence of mitochondrial DNA (mtDNA) has been used to delineate the historical divergence of genetic groups and haplotype frequencies have revealed recent animal movements among local populations (Moritz 1994). MtDNA sequencing is also being increasingly used to investigate the relationships, origins and diversity of domesticated animal species, including rabbits, pigs, goats, buffalos and horses (Kierstein et al. 2004; Kim et al. 1999, 2002; Long et al. 2003; Manjunath et al. 2004; Wang et al. 2000). In contrast, similar studies of aquaculture species are rare (Nguyen et al. 2004), perhaps reflecting their more limited relevance due to the short history of domestication for most species. A notable exception to this is the common carp (Cyprinus carpio L.), the world's oldest domesticated and arguably the most important aquaculture species (Yue et al. 2004).

In China, domestication of common carp commenced over 4000 years ago and nearly three million t are produced annually (FAO 2004). A wide range of molecular marker systems have been used to investigate genetic diversity in carp, including microsatellite (Crooijmans et al. 1997; Yue et al. 2004), randomly amplified polymorphic DNA (RAPDs) (Bartfai et al. 2003), amplified fragment length polymorphism (AFLPs) (David et al. 2001), allozymes (Brody et al. 1979; Kohlmann and Kersten 1999), restriction fragment length polymorphism (RFLPs) (Jian et al. 2003; Kohlmann et al. 2003; Riho et al. 2002) and direct sequencing of mtDNA fragments (Froufe et al. 2002; Wang and Li 2004). However, there have been no comprehensive studies of common carp that utilise nucleotide sequences.

This paper presents the results of sequencing the mtDNA ATPase6/8 and control region fragments for 89 individual common carp representing 32 strains or populations from

around the world. The outcome demonstrates that a genealogical approach shows significant potential for furthering our understanding of the diversity of wild and domesticated carp populations including their origins, genetic history and relationships.

Materials and Methods

Sampling

Tissue samples of common carp were obtained from populations or strains from Vietnam, China, Japan, Indonesia, India, Hungary, Czech Republic, Israel and Australia. Strains, sample codes, and collection details are listed in Table 1.

DNA extraction and amplification of mitochondrial DNA

Total DNA was extracted from fin clip tissue by the protocol described by Crandall et al. (1999). Two to six individual fish were sequenced from

Strain	Sample code 1	Locality	N	Collector
Xingguonensis	XI	Jaing Xi, China	3	Li Sifa
Wananensis	WN		3	
Wuyuanensis	WU		3	
Color	CL		3	
Dor 70	D70	Gan-Shmuel, Israel	4	K.Ilan
Nasice	NA		4	
White Koi	WK		2	U. Lavi
Black Koi	BAK		2	
Kohaku Koi	KK		2	
Sanke Koi	SK		2	
Showa Koi	SHK		2	
Red Koi	КО	Komaki-shi, Japan	2	Y.Lemura
Blatna	CZ	Czech Republic	5	K.Lukas
Hajduboszomeny	HA	Szarvas, Hungary	5	L. Varadi
Szeged	SZ		5	
Tata	TAT		5	
Wild Amur	WA	- Karnataka, India	3	G. C. Mair Y.Basavaraju
Hungarian P3	P3		2	
German X Bangkok	LF		2	
Bhadra River	LB		2	
Majalaya	MA		2	
Rajadanu	RJ	Sukamandi, Indonesia	2	N. Estu
Widan	WI		2	
Rockland Reservoir	AU	Victoria, Australia	2	V. Versace
Yen Bai	YB	Van Chan, Yen Bai, Vietnam	2	v. versace
Tuyen Quang	ТО	Hoang Khai, Tuyen Quang, Vietnam	4	B.T.Thai U.D.Thai
, ,	-	3 , , 3		
Bac Kan	BK	Bach Thong, Bac Kan, Vietnam	4	
Vietnamese white	VN	RIA 1, Bac Ninh, Vietnam	2	
Hungarian scale	H	RIA 1, Bac Ninh, Vietnam	2	
Indonesian yellow	I	RIA 1 , Bac Ninh, Vietnam	4	
GenBanK (X61010)	GB	Taiwan ²	1	Y.Chang H. D. Huang
Goldfish (out group)	GF	Japan	1	M. Murakami

¹ Unique haplotypes within strains are presented by the number 1 and 2 immediately following the sample code (e.g., BK1 and BK2).

each population or strain. The mitochondrial ATPase 6/8 gene fragment was amplified using primers L833 I and H9236 (Lovette et al. 1998) and also the control region of primers designed from common carp sequences on GenBank (AC: X61010),

Carp-Pro (5' AACTCTCACCCTG-GCTACCAAAG 3') and Carp-Phe (5' CTAGGACTCATCTTAG-CATCTTCAGTG 3'). PCR conditions and procedures for visualization, purification and sequencing are given in Murphy and Austin (2002).

Data analysis and phylogenetic reconstruction

Sequences of common carp from GenBank (Chang and Huang 1994) were included in the data set for comparative purposes, and *Carassius*

 $^{^{\}rm 2}$ Location of strain is unknown.

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auratus sequences (GenBank accession number NC 002079) were used as the out group.

The DNA sequences from the control region were aligned using the program Clustal X (Thompson et al. 1997). The DNA sequences from ATPase6/8 regions were aligned by eye. Aligned sequences were imported into PAUP* 4.0.b.10 (Swofford 2000) for phylogenetic analysis. It is important to identify the most suitable model of nucleotide evolution for valid phylogenetic inference (Felsenstein 2004). The Modeltest 3.06 program (Posada and Crandall 1998) was used to test an alternative model of evolution. The best model was then used to calculate pair-wise sequence distances for constructing a neighbor-joining (NJ) tree. Confidence levels in the resulting relationship were assessed using the bootstrap procedure with 1000 replications. Nucleotide (π) and haplotype (h) diversity for specific grouping of samples were calculated using DNASP 4.10 (Rozas et al. 2003).

Results

A total of 820 bp of the ATPase6/8 genes and 745 bp of the control region were sequenced for 87 individuals. All sequences obtained in this study have been submitted to GenBank (accession number: AY597942-AY597985 and AY 600150-AY600241). The mean nucleotide composition is A=31 per cent, T=30 per cent, C=23 per cent and G= 16 per cent. The most suitable evolution model identified for the nucleotide data is: Hasegawa-Kishino-Yano (HKY) + invariable site (I) + gamma distribution shape parameter (G); transition/ transversion ratio = 3.8306. The HKY model allows transition and transversion mutation occurring at different rates and base frequencies of nucleotide to vary (Hasegawa et al. 1985; Page and Holmes 1998). The level of nucleotide

variation at both gene regions was found to be low, although the control region exhibited more variation than the ATPase6/8. Excluding the GenBank Cyprinus carpio (which is of unknown origin) based on nucleotide substitution, the two most divergent ATPase6/8 haplotypes differed by 12 bp and the two most divergent control region haplotypes differed by 14 bp. The average divergence (~ I per cent) is low, given that the control region and ATPase6/8 genes are two of the fastest evolving regions of the mitochondrial genome. The average control region divergence (1.24 per cent) was higher than that of the average ATPase6/8 region (0.68 per cent) (data not shown). Despite this relatively limited

variability, the two gene regions possessed significant phylogenetic signal and were largely consistent in the patterns of variation within and between samples and geographic groupings and the relationships amongst the haplotypes. The nucleotide and haplotype diversity (range 0 to 1) in various geographic groupings is summarized in Figure 1.Although nucleotide diversity is low overall, differences are apparent between the Asian carp (0.008) and carp of European origin (0.000). These differences were most obvious in terms of haplotype diversity (h), which exceeded 0.93 for all carp of Southeast Asian origins but was 0.00 for European carp. Koi carp showed no diversity, were identical

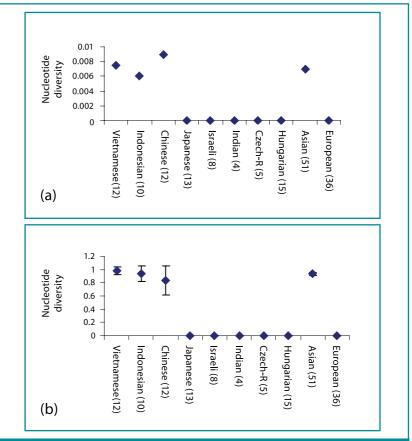


Figure 1.(a) & (b). Nucleotide and haplotype diversity of geographic common carp group derived from combination ATPase6/8 and control regions. The number in bracket is number of individual sequenced. Japanese carp sample (Koi carp) were obtained from Japan and Israel. Asian carp include: Vietnamese, Chinese, Japanese, Indian and Indonesian carp. European group include samples from Israel, Australia, India, Vietnam, Czech Republic (Czech-R) and Hungarian.

to the Chinese Color (CL) carp and showed minimal divergence from Vietnamese carp. The European carp, while clearly distinguishable from all other carp, are closely related to the clade containing the Koi and Vietnamese carp. Significant haplotype diversity is apparent among the Indonesian and Chinese samples. The most divergent haplotype was represented by the Chinese Wuyuanensis strain (WU) that differs from between 6 to 20 bp from all other common carp haplotypes. In addition, the sample of Amur river carp (WAI and WA2) is interesting as it contains two haplotypes, one close to Chinese Color carp and therefore also Vietnamese carp, and the other that is identical to European carp. The relationship of the carp strains is shown in Figure 2.

Discussion

Mitochondrial variation

This study has demonstrated that direct sequencing of variable mtDNA fragments can provide useful insights into the genetic diversity, origin, divergence and genealogy of carp strains and populations.A surprising result was the low levels of divergence observed in what are generally considered to be the most rapidly evolving vertebrate mtDNA fragments, the ATPase6/8 genes and the control region (Verspoor 1998), especially for a freshwater species with such an extensive distribution across Eurasia. A second surprising result was the complete absence of variation within two major strains of carp, the European common carp and Koi carp. These findings are consistent with the long history of domestication of this species which has undoubtedly involved significant founder and population bottleneck events leading to localized loss of genetic variation (Balon 1995).

Genetic relationships

The origin of a number of carp strains is uncertain (Balon 1995). Koi carp for example, are thought to have been developed directly from common carp stocks in Japan, or Chinese Color carp or German pond carp (Balon 1995; Wang and Li 2004). The results of this study suggest that Koi carp have originated from Chinese Color carp which have a long history of domestication that can be traced back over 1200 years (Wang and Li 2004). This is consistent with the results of Wang and Li (2004) based on COII sequences, but contrary to those of Froufe et al. (2002) who sequenced 540 bp of the control region and found that their single sample of Japanese Koi (N=4) had a haplotype identical to European carp. This anomaly could be due to mixing of stocks with Koi carp acquirng the European mtDNA through cross breeding. Despite their substantial color polymorphisms, only a single haplotype was detected among the 12 individual Koi carp (representing 6 strains) sequenced. This result is consistent with Koi and Chinese Color carp having passed through a substantial bottleneck during the domestication process.

Carp from Europe or originating from Europe had the same haplotype, which was divergent from all other haplotypes detected. A remarkable feature of the European carp samples is the complete absence of any variation, which suggests a history of founder effects and small population size associated with translocation and domestication, like the Koi carp. In contrast, Chinese carp showed considerable haplotype diversity and divergence. While no differences were detected between the Xingguonensis (XI) and Wananensis (WN) strains, their haplotypes are highly differentiated from Chinese Color carp (CL) and Wuyuanensis (WU) carp with the latter showing

the most distinctive common carp haplotype found so far. This suggests that carp have a longer evolutionary history in China, compared to other countries or geographic regions (i.e., Vietnam and Europe).

The genetic makeup of Amur river carp represents an anomaly in that, while they occur over 5000 km from northern Vietnam and 10 000 km from Central Europe, they possess one haplotype that differs by only two bases from a haplotype found at Tuyen Quang in northern Vietnam and another haplotype identical to the European carp. As the Amur river carp examined were originally derived from a live gene bank line in the Fish Culture Research Institute Szarvas, Hungary, presently maintained at Karnataka in India, this finding might be attributed to accidental stock mixing in captivity. However, Froufe et al. (2002) found a similar result for a sample of Amur river carp obtained from the wild, with some haplotypes being very close to European carp and others to Asian carp. They interpreted this finding to indicate an Asian origin for European carp. However, the converse possibility, that European carp had been introduced to China, has also been proposed (Balon 1995) and is also consistent with the data.

Taxonomic implications

The taxonomy of common carp is highly contentious and confusing with over 30 synonyms of *C. carpio* listed on Fish-Base (www.fishbase. org/search.cfm) and a proliferation of named strains. Considering only relatively recent literature, Kirpichnikov (1967) recognised four subspecies: *Cyprinus carpio carpio* (Europe); *C. c. aralensis* (Central Asia); *C. c. haematopterus* (East Asia); and *C. c. viridiviolaceus*. (Southeast Asia). In contrast, Balon (1995) recognized only two subspecies: *Cyprinus carpio carpio* (Europe) and

Cyprinus carpio haematopterus (East Asia). Subsequently, Kirpichnikov (1999) questioned the validity of C. c. viridiviolaceus whereas Li et al. (2001) recognised four morphologically distinctive red common carp strains from China which he referred to as C. c. xinggguonensis, C. c. wannanensis, C. c. wuyanensis, C. c. color. In one of the most recent taxonomic treatments of common carp Kottelat (2001) considers the common carp cultured in Southeast Asia to be a distinct species, C. rubrofuscus, although this is disputed by Nguyen and Ngo (2001) who consider this species to be quite rare. The results of genetic studies using allozyme, microsatellite, and RFLP analysis of the mtDNA ND-3/4 and ND-5/6 regions by Kohlman et al. (2003) were interpreted to provide support for Balon's (1995) position for the existence of two subspecies, C. c. carpio and C. c. haematopterus, as the Asian and European samples form two distinct clades.

MtDNA sequence data, often used for clarifying species boundaries (Avise 2000), provides little support for any of the previous positions. While the results of this study are largely consistent with those of Kohlman et al. (2003) with regard to the distinctiveness of the European carp from Vietnamese/Amur carp/Koi carp, the finding that samples of carp from China or of Chinese origin are highly divergent fails to support an independent origin for western and eastern common carp lineages and their recognition as distinct subspecies (or species). Indeed, this finding, combined with the deepest branches of the phylogenetic tree represented by samples of Chinese origin, suggests that common carp most probably originated in China.

While genetic distance is not always a reliable guide to taxonomic recognition, the levels of divergence observed for the control and ATPase6/8 regions (~I per cent) are

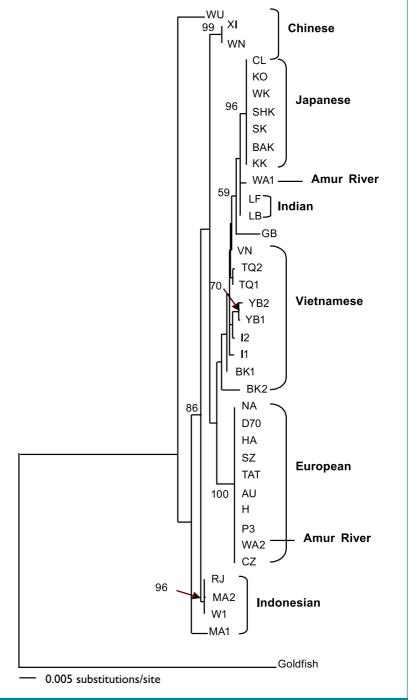


Figure 2. Neighbour-joining tree derived from ATPase6/8 and control region, using HKY+I+G model of evolution. Bootstrap values are base on 1000 replicates. Bootstrap value is given for nodes with at least 50% or more support.

much less in carp compared to that usually observed among fish species and sub-species. For example, Hurt et al. (2001) observed between 15-18 per cent divergences in control

region sequences among closely related species within the genus Acanthopagrus. It is essential that some stability to the taxonomy of the common carp is achieved and

we recommended that this study be extended to include more samples of carp, especially from China, with parallel studies of morphology, genetic variation at rapidly evolving nuclear loci (especially intron regions) and reproductive relationships.

The Development of a Global Genealogy of Common Carp Strains

The findings of this study indicate that the establishment of genealogical relations among carp strains based on mtDNA sequencing can provide new and useful information regarding the history and development of carp strains. For example, this study has shown that Koi, Chinese, Indonesian yellow and Vietnamese white carp are all closely related and that the other Indonesian and some of the Chinese common carp are highly divergent (e.g., Wuyuanensis). Further, this approach can reveal carp stocks of mixed origin (e.g., Majalaya) and others that appear to have been through genetic bottlenecks (e.g., Koi and European carp). Information on relationships among strains can be used as a guide to the effective mixing of strains as part of genetic improvement programs (Ferguson 1995). It can also provide information on the historical origins of strains, whether they represent a composite of different strains, and if they have lost variation during the process of domestication.

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References

- Avise, J.C. 2000. Phylogeography- The History and Formation of Species. Harvard University Press, USA.
- Balon, E.K. 1995. Origin and domestication of wild carp, Cyprinus carpio: from Roman gourmets to the swimming flowers. Aquaculture 129:3-48.
- Bartfai, R., S. Egedi, G.H. Yue, B. Kovacs, B. Urbanyi, G. Tamas, L. Horvath and L. Orban. 2003. Genetic analysis of two common carp broodstocks by RAPD and microsatellite markers. Aquaculture 219:157-167.
- Brody, T., D. Kirsht, G. Parag, G.
 Wohlfarth, G. Hulata and R. Moav.
 1979. Biochemical genetic comparison of the Chinese and European races of the common carp.
 Animal Blood Group Biochemical.
 Genetic 10:141-149.
- Chang, Y., and H.D. Huang. 1994. The complete nucleotide sequence and gene organisation of carp (Cyprinus carpio) mitochondrial genome. Journal of Molecular Evolution 38:138-155.
- Crandall, K.A., J.S.H. Fetzner, M. Lawler, M. Kinnersly and C.M. Austin. 1999. Phylogenetic relationships among Australian and New Zealand genera of freshwater crayfish. Australian Journal of Zoology 47:199-214.
- Crooijmans, R.P.M., V. Bierbooms, J. Komen, J.J. Vanderpoel and M. Groenen. 1997. Microsatellite markers in common carp (*Cyprinus carpio L.*). Animal Genetics 28:129-134.
- David, L., P. Rajasekaran, J. Fang, J. Hillel

- and U. Lavi. 2001. Polymorphism in ornamental and common carp strains (Cyprinus carpio L.) as revealed by AFLP analysis and new set of microsatellite markers. Molecular Genetic Genomics 266:353-362.
- FAO. 2004. Common carp production. http://www.fishbase.org/report/FAO/FAOAquacultureList.cfm?scientific=Cyprinus%20carpio.
- Felsenstein, J. 2004. Inferring phylogenies. Sinaner Associates, Massachusetts (664).
- Ferguson, M.M. 1995. The role of molecular genetic markers in the management of cultured fishes, p. 81-104. In: G.R. Carvalho and T.J. Pitcher (eds.) Molecular Genetics in Fisheries. Chapman & Hall, Great Britain.
- Froufe, E., I. Magyary, I. Lehoczky and S. Weiss. 2002. mtDNA sequence data supports an Asian ancestry and single introduction of common carp into Danube Basin. Fish Biology 61:301-304.
- Hasegawa, M., H. Kishino and T.A. Yano. 1985. Dating of the humanape splitting by a molecular clock of mitochondrial DNA. Journal of Molecular Evolution 22:160-174.
- Hurt, A.C., N.P. Murphy, J.G. Patil and C.M. Austin. 2001. Molecular genetic evidence for a new species of bream of genus Acanthopagrus Peter (*Perciformes: Sparidae*). Asian Fisheries Science 14:425-433.
- Jian, F.Z., J.W. Quing, Z.Y.Yu and G.T. Jin. 2003. Genetic divergence between Cyprinus carpio carpio and Cyprinus carpio haematopterus as assessed by mitochondrial DNA analysis, with emphasis on origin of European domestic carp. Genetica 119:93-97.
- Kierstein, G., M. Vallinoto, A. Silva, M.P. Schneider, L. lannuzzi and B. Brenig. 2004. Analysis of mitochondrial D-loop region casts new light on domestic water buffalo (Bubalus bubalis) phylogeny. Molecular Phylogenetics and Evolution 30:308-324.
- Kim, K.I., J.H. Lee, K. Li, Y.-P. Zhang,

- S.-S. Lee, J. Gongora and C. Moran. 2002. Phylogenetic relationships of Asian and European pig breeds determined by mitochondrial DNA D-loop sequence polymorphism. Animal Genetics 33:19-25.
- Kim, K.I., Y.H. Yang, S.S. Lee, C. Park, R. Ma, J.L. Bouzat and H.A. Lewin. 1999. Phylogenetic relationships of Cheju horses to other horse breeds as determined by mtDNA D-loop sequence polymorphism. Animal Genetics 30:102-108.
- Kim, S.K., S.J. Yeo and B.C. Choi. 2002. Genetic diversity of north-east Asian cattle based on microsatellite data. Animal Genetics 33:201-204.
- Kirpichnikov, V.S. 1967. Homologous hereditary variation and evolution of wild common carp *Cyprinus carpio L*. Genetika 8:65-72 (in Russian).
- Kirpitchnikov, V.S. 1999. Genetics and breeding of common carp. INRA, Pari, France.
- Kohlmann, K., R. Gross, A. Murakaeva and P. Kersten. 2003. Genetic variability and structure of common carp (Cyprinus carpio) populations throughout the distribution range inferred from allozyme, microsatellite and mitochondrial DNA markers. Aquaculture Living Resources 16:421-431.
- Kohlmann, K., and P. Kersten. 1999. Genetic variability of German and foreign common carp (Cyprinus carpio L.) populations. Aquaculture 173:435-445.
- Kottelat, M. 2001. Fishes of Laos. WHT Publication (Pte) Ltd., 95, Cotta, Colombo 5, Sri Lanka.
- Li, S., C.Wanqi, Z. Shuming and W. Chenghui. 2001. Genetic research program and planned activities in Shanghai Fisheries University (SFU), China. Report submitted to the Sixth Steering Committee Meeting of International Network on Genetics in Aquaculture (INGA), 8-10 May 2001, Hanoi, Vietnam
- Long, J.R., X.P. Qiu, F.T. Zeng, L.M. Tang and Y.P. Zhang. 2003. Origin of

- the rabbit (Oryctolagus cuniculus) in China: evidence from mito-chondrial DNA control region sequence analysis. Animal Genetics 34:82-87.
- Lovette, I. J., E. Bermingham, G. Seutin and R. Ricklest. 1998. Evolutionary differentiation in three endemic West Indian warblers. Auk 115:890-903.
- Manjunath, B.J., K.R. Pramod, K.M. Ajoy, T.S. Chris, S. Lalji and T. Kumarasamy. 2004. Phylogeography and origin of Indian domestic goats. Molecular Biology and Evolution 21:454-462.
- Moritz, C. 1994. Applications of mitochondrial DNA analysis in conservation: a critical review. Molecular Ecology 3:401-411.
- Murphy, N.P. and C.M. Austin. 2002.

 Molecular taxonomy of Australian Macrobrachium shrimps
 (Palaemonidae: Decapoda) reveals
 anomalies in their current classification. Invertebrate Systematics
 16:697-701.
- Nguyen, T.T.T., C.M.Austin, M.M. Meewan, M.B. Schultz and D.R. Jerry. 2004. Phylogeography of the freshwater crayfish Cherax destructor Clark in inland Australia: Historical fragmentation and recent range expansion. Biological Journal of the Linnean Society 81:539-550.
- Nguyen, V.H. and S.V. Ngo. 2001. Vietnamese freshwater fish. Agriculture Publish House, Hanoi. (in Vietnamese).
- Page, M.D.R., and C.E. Holmes. 1998. Molecular Evolution: A Phylogenetic Approach. Blackwell Science Ltd., Great Britain.
- Posada, D., and A.K. Crandall. 1998. Modeltest: testing the model of DNA substitution. Bioinformatics 14:817-818.
- Riho, G., K. Klaus and K. Petra. 2002. PCR- RFLP analysis of the mitochondrial ND- 3/4 and ND- 5/6 gene polymorphism in the European and East Asian subspecies of common carp (Cyprinus carpio L). Aquaculture 204:507-516.
- Rozas, J., J.C. Sanchez-Delbarrio, X.

- Messeguer and R. Rozas. 2003. Dna SP, DNA Polymorphism analyses by coalescent and other methods. Bioinformatics 19:2496-2497
- Swofford, D.L. 2000. PAUP* Phylogenetic Analysis Using Parsimony (*and Other Methods). 4. Sinauer Associates: Sunderland, Massachusetts
- Thompson, J.D., T.J. Gibson, F. Plewniak, F. Jeanmougin and D.G. Higgens. 1997. The clustal X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. Acids. Res. 24:4876-4882.
- Verspoor, E. 1998. Molecular markers and genetic management of farmed fish, p. 355-377 In: D.K. Black and D.A. Pickering (eds). Biology of Farmed Fish. Sheffield Academic Press, England.
- Wang, C.H. and S.F. Li. 2004. Phylogenetic relationships of ornamental (koi) carp, Oujiang color carp and long-fin carp revealed by mitochondrial DNA COII gene sequences and RAPD analysis. Aquaculture 231:83-91.
- Wang, Z., P. Jayasankar, K.S. Khoo, K.
 Nakamura, K. Sumantadinata, O.
 Carman and N. Okamoto. 2000.
 AFLP fingerprinting reveals genetic variability in common carp stock from Indonesia. Asian Fisheries
 Science 13:139-147.
- Yue, G.H., M.Y. Ho, L. Orban and J. Komen. 2004. Microsatellites within genes and ESTs of common carp and their applicability in silver crucian carp. Aquaculture 234:85-98.

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